Facile and Stereospecific Synthesis of (S)- and (R)-[2-²H]lsopentenyl Pyrophosphates ¹

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(S)- and (R)- $[2-^{2}H]$ lsopentenyl[†] pyrophosphates, which are useful precursors for stereochemical studies on the biosynthesis of isoprenoids, were chemically synthesized in five steps *via* (S)- and (R)-3-methyl-2,3-epoxybutan-1-ols starting with 3,3-dimethylallyl alcohol, in 18% and 15% overall yields, respectively.

During the course of our work on the unusual elimination of the pro-4S hydrogen atom of mevalonic acid (MVA) in the process of the Z-prenyl chain elongation in the biosynthesis of the polyprenols by higher plants,² we required (S)- and (R)- $[2^{-2}H]$ -

$$\int_{R^{1}}^{5} \frac{1}{R^{2}} H_{2}OR^{3}$$
(1) $R^{1} = H, R^{2} = {}^{2}H, R^{3} = P_{2}O_{6}(NH_{4})_{3}$
(2) $R^{1} = {}^{2}H, R^{2} = H, R^{3} = P_{2}O_{6}(NH_{4})_{3}$
(7) $R^{1} = H, R^{2} = {}^{2}H, R^{3} = Ts$
(11) $R^{1} = {}^{2}H, R^{2} = H, R^{3} = Ts$
(12) $H_{1} = {}^{2}H, R^{2} = H, R^{3} = Ts$
(13) $H_{1} = {}^{2}H, R^{2} = H, R^{3} = Ts$
(14) $H_{1} = {}^{2}H, R^{2} = H, R^{3} = Ts$
(15) $H_{2}OH$
(16) (17) $H_{1} = {}^{2}H, R^{3} = Ts$
(17) $H_{1} = {}^{2}H, R^{3} = Ts$
(18) $H_{1} = {}^{2}H, R^{3} = Ts$
(19) $H_{1} = {}^{2}H, R^{3} = Ts$
(19) $H_{1} = {}^{2}H, R^{3} = Ts$
(11) $H_{1} = {}^{2}H, R^{3} = H, R^{3} = Ts$
(11) $H_{1} = {}^{2}H, R^{3} = Ts$
(12) $H_{2}OH$
(13) $H_{1} = {}^{2}H, R^{3} = Ts$
(14) $H_{2} = H, R^{3} = Ts$
(15) $H_{2}OH$
(16) $H_{2}OH$
(17) $H_{2}OH$
(18) $H_{2} = H, R^{3} = Ts$
(19) $H_{2} = H, R^{3} = Ts$
(19) $H_{2} = H, R^{3} = Ts$
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(13) $H_{1} = {}^{2}H, R^{3} = Ts$
(14) $H_{2} = H, R^{3} = Ts$
(15) $H_{2} = H, R^{3} = Ts$
(17) $H_{2} = H, R^{3} = Ts$
(18) $H_{2} = H, R^{3} = Ts$
(19) H_{2

(2) has yet been developed, although (S)- and (R)- $[2-^{3}H]$ IPPs have been enzymically synthesized from (4R)- and (4S)- $[4-^{3}H]$ -mevalonic acids (MVAs), respectively.³⁻⁵ Previous attempts to synthesize compounds (1) and (2) in multigram quantities failed

(3)
$$R = H$$

(12) $R = Ts$
(15) $R = P_{2}O_{c}(NH_{c})_{2}$

E



(5) $R^{1} = H$, $R^{2} = {}^{2}H$, $R^{3} = CH_{2}OH$ (6) $R^{1} = H$, $R^{2} = {}^{2}H$, $R^{3} = CH_{2}OTs$ (9) $R^{1} = {}^{2}H$, $R^{2} = H$, $R^{3} = CH_{2}OH$ (10) $R^{1} = {}^{2}H$, $R^{2} = H$, $R^{3} = CH_{2}OTs$ (13) $R^{1} = H$, $R^{2} = {}^{2}H$, $R^{3} = CHO$ (14) $R^{1} = {}^{2}H$, $R^{2} = H$, $R^{3} = CHO$



isopentenyl pyrophosphates (IPPs) (1) and (2) for elucidation of the stereochemistry of hydrogen elimination in the enzymic formation of E- and Z-prenyl chains of the polyprenols. No chemical synthesis of stereospecifically deuteriated IPPs (1) and

(8)

† In this paper isopentenyl refers to the 3-methylbut-3-enyl radical.

due to troublesome procedures for the preparation of the enzyme system and the synthesis of stereospecifically deuteriated MVA. We therefore developed a facile and stereospecific chemical synthesis of compounds (1) and (2), and here describe the results.

Compounds (1) and (2) were synthesized from 3,3-dimethylallyl alcohol (3) by a five-step procedure; (i) asymmetric epoxi-



Figure. Circular dichroism curves of compounds (13) and (14) in diethyl ether

dation of compound (3), (ii) epoxide opening with $LiAl^2H_4$, (iii) selective toluene-p-sulphonylation, (iv) regiospecific dehydration, and (v) pyrophosphorylation, as follows. Following the method of Sharpless,⁶ epoxidation of 3,3-dimethylallyl alcohol (3) using (+)-diethyl tartrate as a chiral source yielded (S)-3-methyl-2,3-epoxybutan-1-ol (4) (55% yield after distillation). The optical purity of the epoxide (4) was found to be in >89% enantiomeric excess (e.e.) by comparison with literature values.⁷ Reduction of the epoxide (4) with $LiAl^2H_4$ gave (R)-[2-²H]-3-methylbutane-1,3-diol (5) in 82% yield (99% incorporation of 2 H). Toluene-*p*-sulphonylation of the diol (5) with toluene-p-sulphonyl chloride in dry pyridine gave the (R)monotoluene-p-sulphonate (6) in 68% yield which was dehydrated with methanesulphonyl chloride and triethylamine to give the (S)-isopentenyl toluene-p-sulphonate (7) in 71%yield. Pyrophosphorylation⁸ of the compound (7) gave (S)- $[2-^{2}H]$ IPP (1) in 83% yield. The overall yield of compound (1) from compound (3) was 18%.

In the preparation of (R)- $[2-^{2}H]IPP$ (2), (R)-3-methyl-2,3epoxybutan-1-ol (8) (>88% e.e.) was obtained from compound (3) by Sharpless oxidation using unnatural (-)-diethyl tartrate as a chiral source. The epoxy alcohol (8) was converted into (R)- $[2-^{2}H]IPP$ (2) via compounds (9), (10), and (11), successively, in a manner similar to that described above. The overall yield of compound (2) from compound (3) was 15%.

Although the dehydrated products derived from compounds (6) and (10) were carefully analysed by means of h.p.l.c. (Radial Pak B10) and t.l.c. (Merck precoated silica gel $60F_{254}$ plate), the internal alkene, 3,3-dimethylallyl toluene-*p*-sulphonate (12), was not detected. Furthermore, the ¹H n.m.r. spectra of the dehydrated products exhibited no signals due to the internal alkene (12), thus confirming that the dehydration of compounds (6) and (10) forms specifically the isopentenyl toluene-*p*-sulphonates (7) and (11), respectively.

The absolute configuration at C-2 of the diol (5) was determined on the basis of a positive Cotton effect in the circular dichroism spectrum of the aldehyde (13), which was obtained from the diol (5) by oxidation with pyridinium chlorochromate. The positive Cotton effect associated with the $n \rightarrow \pi^*$ transition of the carbonyl group is shown in the Figure (a). On the other hand, the aldehyde (14) from the diol (9) derived from the epoxide (8) showed a negative Cotton effect in its c.d. spectrum, as shown in the Figure (b). It has been shown that (R)- α -deuteriated aldehydes show a negative Cotton effect,⁹ thus, the absolute configurations of the aldehydes (13) and (14) were determined to be S and R, respectively. Accordingly, the absolute configurations of the deuteriated IPPs (1) and (2) should be S and R, respectively.

The absolute configurations of (1) and (2) were further examined by feeding experiments using pig liver farnesyl pyrophosphate (FPP) synthetase, as follows. When compound (1) and dimethylallyl pyrophosphate (DMAPP) (15) were used as precursors, (E,E)-farnesol (16), thus enzymically synthesized, exhibited the molecular ion peak at m/z 224; this clearly indicates the retention of the deuterium atom of compound (1) in the biological formation of (E,E)-farnesol. When compound (2) and DMAPP (15) were used as precursors, on the other hand, the mass spectrum of the enzymically synthesized (E,E)-farnesol (17) was the same as that of authentic farnesol, indicating the loss of the deuterium atom of compound (2). According to Cornforth's stereochemical picture for the biosynthesis of isoprenoids, $^{10-12}$ the pro-2S hydrogen of IPP is retained and the pro-2R hydrogen is eliminated during the formation of E-prenyl chain in the biosynthesis of farnesol with the pig liver enzyme, thus the absolute configurations assigned to (1) and (2) as described above were established to be correct.

This synthetic method is favourable for the large-scale preparation of stereospecifically labelled IPP, because the sequence is simple and the reaction time at each step is relatively short. In addition, $LiAl^{3}H_{4}$, instead of the deuteride can be used in the sequence to yield the stereospecifically tritiated IPP.

Experimental

¹H and ¹³C N.m.r. spectra were recorded on Hitachi R-600 FT n.m.r. (60 MHz) and Hitachi R-42 FT n.m.r. spectrometers, respectively; chemical shifts are given in δ (p.p.m.) downfield from internal SiMe4 and [2,3-2H4]-3-(trimethylsilyl)propionic acid sodium salt (TSP) for a [2H]chloroform solution and a ²H₂O solution, respectively. Electron impact (e.i.) mass spectra were obtained on a Shimadzu QP-1000 spectrometer operating with an ionization potential of 70 eV. Chemical ionization (c.i.) mass spectra were recorded on a Shimadzu QP-1000 spectrometer using isobutane as the carrier gas. Optical rotation was measured with a JASCO DIP-360 digital polarimeter, using a 1.0-dm cell. C.d. spectra were recorded on a JASCO J-40 spectropolarimeter. G.l.c. was carried out on a 2 m column packed with 2% OV-17 or 15% DEGS on Chromosorb W (AW-DMCS; 80-100 mesh) with N₂ gas. H.p.l.c. analysis was performed on a Waters Radial-pak B10 column with a flow rate of 0.6 ml min⁻¹ and the column effluent was monitored at 261 nm.

(S)-3-Methyl-2,3-epoxybutan-1-ol (4).—Titanium tetraisopropoxide (11.9 ml, 40 mmol), (+)-diethyl tartrate (7.0 ml, 40 mmol), 3,3-dimethylallyl alcohol (3) (3.4 g, 40 mmol), and tbutanoic acid (3.0M) in toluene (Aldrich) (26.9 ml, 80 mmol) was added to dichloromethane (400 ml) at -20 °C (solid CO₂-CCl₄ bath), successively, and the mixture was maintained at the same temperature for 24 h. After addition of Me₂S (9.9 g), the mixture was stirred for 45 min at -20 °C and poured into saturated NaF solution (800 ml) with vigorous stirring. After having been stirred for 14 h, the mixture was saturated with NaCl and filtered through a Celite pad. The aqueous layer was extracted with dichloromethane (×3), the combined organic phase was dried over Na₂SO₄ and concentrated to give an oily residue (9.5 g). This residue was distilled under reduced pressure to give the epoxy alcohol (4) (2.22 g, 55%) as a colourless oil: b.p. 58—60 °C (7 mmHg) and $[\alpha]_D^{25} - 20.1 \pm 0.9^\circ$ (*c* 0.42 in CHCl₃) {lit.,⁷ $[\alpha]_D^{25} + 21.0^\circ$ (*c* 1.71 in CHCl₃) for the *R*-isomer; >93% e.e.}; n_D^{25} 1.427; d_4^{25} 0.9912; 99.8% pure on g.l.c.; v_{max} .(neat) 3 420 (OH) and 1 250 cm⁻¹ (epoxy); $\delta_{\rm H}$ (CDCl₃) 1.32 and 1.35 (each 3 H, s, 2 × Me), 2.54 (1 H, br t, OH, disappeared in D₂O), 2.88—3.06 (1 H, m, 2-H), and 3.57—4.02 (2 H, m, 1-H); *m/z* (e.i.) 85 (5%) and 59 (100); *m/z* (c.i.) 103 (*M*⁺ + H, 16%), 102 (*M*⁺, 12), and 85 (100).

 (\mathbf{R}) - $[2-^{2}\mathbf{H}]$ -3-Methylbutane-1,3-diol (5).—A solution of epoxide (4) (6.13 g, 60 mmol) in dry diethyl ether (50 ml) was added dropwise to a stirred suspension of $LiAl^2H_4$ (2.0 g) in dry diethyl ether. The mixture was refluxed for 16 h, treated with saturated Na_2SO_4 solution (5 ml), and stirred for a further 1 h. The white solid was filtered off and the filtrate was dried over Na_2SO_4 ; the white solid was then extracted with a 20 ml portion of CHCl₃ (\times 3). The combined organic phase was concentrated to give the diol (5) (5.17 g, 82%) as a colourless oil; $[\alpha]_D^{25}$ $-0.1 \pm 0.1^{\circ}$ (c 3.55 in CHCl₃); $n_{\rm D}^{25}$ 1.436; d_4^{25} 0.9923; 99.8% pure on g.l.c.; v_{max} (neat) 3 370 (OH) and 2 170 cm⁻¹ (CD); $\delta_{\rm H}(\rm CDCl_3)$ 1.27 (6 H, s, 2 × Me), 1.70 (1 H, m, 2-H), 3.65 (2 H, br, $2 \times OH$, disappeared in D₂O), and 3.85 (2 H, d, J 6 Hz, 1-H); m/z (e.i.) 90 (M^+ – Me, 10%), 72 (18), 59 (95), and 43 (100); m/z (c.i.) 106 (M^+ + H, 4%), 105 (M^+ , 2), 88 (M^+ - H₂O + H, 65), and 70 $(M^+ - 2H_2O + H, 100)$. The deuterium content was determined to be 99% by e.i.m.s. spectrometry.

Oxidation of (R)-[2-²H]-3-Methylbutane-1,3-diol (5).-Compound (5) (110 mg) in dry dichloromethane (2 ml) was added to a slurry of pyridinium chlorochromate (340 mg) in dry dichloromethane (2 ml), and the mixture was stirred at room temperature for 3 h. Dry diethyl ether (10 ml) was then added to the reaction mixture, and the resulting slurry was stirred for another hour. The slurry was passed through a column packed with 5 g of Florisil, and the eluate was concentrated under reduced pressure. The condensate was purified on a silica gel column with diethyl ether-hexane (1:1) to give (S)-[2-²H]-3hydroxy-3-methylbutanal (13) (43 mg, 40%) as a colourless oil; v_{max} (neat) 3 400 (OH), 2 180 (CD), and 1 708 cm⁻¹ (C=O); $\delta_{\rm H}({\rm CDCl}_3)$ 1.34 (6 H, s, 2 × Me), 2.60 (1 H, d, J 2 Hz, 2-H), and 9.83 (1 H, d, J 2 Hz, 1-H); m/z (e.i.) 88 (M^+ – Me, 8%), 59 (49), 57 (21), and 43 (100); m/z (c.i.) 104 (M^+ + H, 4%) and 86 $(M^+ - H_2O + H, 100)$. This methylbutanal was immediately used for c.d. measurements.

(**R**)-[2-²H]-3-*Hydroxy*-3-*methylbutyl* Toluene-p-sulphonate (6).—A solution of toluene-p-sulphonyl chloride (14.1 g, 74 mmol) in dry pyridine (5 ml) was added dropwise to a stirred solution of compound (5) (5.17 g, 49 mmol) in the same solvent (5 ml) at 0 °C under a nitrogen atmosphere. The solution was stirred at 0 °C for 20 min and left at 4 °C for 2 days, and then poured onto ice. The resulting mixture was extracted with ether (× 2), and the combined organic phase washed successively with cold 5% HCl, 5% NaHCO₃, and saturated NaCl solutions and then dried (Na₂SO₄). Evaporation of the solvent under reduced pressure gave a colourless oil (9.86 g). This oil was chromatographed on a silica gel column, using EtOAc-hexane (1:4) as eluant to give the pure monotoluene-p-sulphonate (6) (8.70 g, 68%) as a colourless oil; $[\alpha]_D^{25} - 0.13 \pm 0.02^\circ$ (neat); n_D^{25} 1.504; d_4^{25} 1.1684; λ_{max} .(EtOH) 273 (log ε 2.38), 267 (2.46), 261 (2.51), 255 (2.48), and 249 nm (2.31); v_{max} .(neat) 3 540—3 420 (OH), 2 170 (CD), 1 598, and 1 495 cm⁻¹ (Ar); δ_H (CDCl₃) 1.21 (6 H, s, 2 × Me), 1.74 (1 H, br s, OH, disappeared in D_2O), 1.83 (1 H, m, 2-H), 2.45 (3 H, s, ArMe), 4.21 (2 H, d, J 7 Hz, 1-H), and 7.27—7.87 (4 H, m, ArH); m/z (e.i.) 259 (M^+ , 0.1%), 244 ($M^+ -$ Me, 9), 230 (5), 172 (69), 155 (21), 91 (66), 72 (82), 60 (100), and 45 (78); m/z (c.i.) 260 ($M^+ +$ H, 4%) and 242 ($M^+ -$ H₂O + H, 100). The e.i.m.s. spectrum indicated that little or no deuterium was lost during the toluene-*p*-sulphonylation of (5). The monotoluene-*p*-sulphonate (6) showed a single peak on analytical h.p.l.c. which was eluted with EtOAc-hexane (1:9).

(S)-[2-²H]-3-Methylbut-3-enyl Toluene-p-sulphonate (7) (Isopentenyl Toluene-p-sulphonate).—A solution of compound (6) (3.21 g) in dry ether (100 ml) containing methanesulphonyl chloride (3.0 g) was cooled to 0 °C. Triethylamine (20 g) was added dropwise to the solution over 1 h with vigorous stirring at 0 °C. Stirring was continued at room temperature for 1 h. The mixture was filtered and the filtrate was evaporated to dryness. The crude product was purified by silica gel chromatography with EtOAc-hexane (1:9) as eluant to give the title compound (7) (2.12 g, 71%) as a colourless oil; $[\alpha]_D^{25} - 0.47 \pm 0.03^\circ$ (c 13.8 in CHCl₃); n_D^{25} 1.511; d_4^{25} 1.1304; λ_{max} .(EtOH) 273 (log ε 2.39), 267 (2.47), 261 (2.52), 255 (2.49), and 249 nm (2.30); v_{max}(neat) 3 090, 1 650, and 898 (C=CH₂), 2 150 (CD), 1 600, and 1 495 cm⁻¹ (Ar); δ_{H} (CDCl₃) 1.66 (3 H, s, Me), 2.31 (1 H, m, 2-H), 2.45 (3 H, s, ArMe), 4.14 (2 H, d, J 7 Hz, 1-H), 4.68 and 4.79 (2 H, m, 4-H), and 7.27-7.88 (4 H, m, ArH); m/z (e.i.) 155 (17%), 91 (59), 69 (100), and 68 (61); m/z (c.i.) 242 (M^+ + H, 15%) and 89 (100). On analytical h.p.l.c. the isopentenyl toluene-p-sulphonate (7) which was eluted with EtOAc-hexane (1:19), diethyl ether-hexane (1:9), and CH_2Cl_2 -hexane (1:19)showed a single peak.

(S)-[2-²H]-3-Methylbut-3-enyl Pyrophosphate (1).—Pyrophosphorylation of compound (7) (1.10 g, 4.6 mmol) with tristetrabutylammonium hydrogen pyrophosphate (12.1 g) was carried out following the method reported previously.8 Cellulose column chromatography of the reaction mixture gave the pure pyrophosphate (1) (1.128 g, 83%) as a highly hygroscopic solid; $[\alpha]_D^{25} - 0.19 \pm 0.05^\circ$ (c 8.18 in H₂O); v_{max} (KBr) 3 600-2 600 (NH), 2 180 (CD), 1 645 (C=C), and 1 195 cm⁻¹ (P=O); $\delta_{H}(D_{2}O)$ 1.79 (3 H, s, Me), 2.38 (1 H, t, J 7 Hz, 2-H), and 4.06 (2 H, t, J 6.5 Hz, 1-H). The signals of the terminal alkene protons, expected to be present at $ca. \delta_{H} 4.8$, overlapped with the solvent signal (δ_H 4.75–4.90); $\delta_C(D_2O)$ 146.9 (C-3), 114.7 (C-4), 67.5 (d, J_{13</sup>_{C,P} 3 Hz, C-1), 40.6 (dd, J₁₃_{C,2H} 20 Hz, J₁₃_{C,P} 7 Hz,} C-2), and 24.7 (C-5). The IPP (1) showed a single spot on analytical t.l.c. (Merck precoated cellulose plate). The spot was visualized according to the procedures reported previously.8

Feeding Experiments.—A mixture of MgCl₂ (5 mmol), 1,4dithiothreitol (4 mmol), KF (5 mmol), iodoacetamide (5 mmol), compound (15) (170 μ mol), compound (1) or (2) (170 μ mol), and pig liver farnesyl-pyrophosphate synthetase ¹³ (100 mg) in Tris-HCl buffer solution (20 mM, 500 ml; pH 7.7) was incubated at 37 °C for 24 h and then treated with alkaline phosphatase as usual. The reaction mixture was extracted with hexane, and the extract subjected to g.l.c.–m.s. analysis. The farnesols (16) and (17) synthesized enzymically were estimated at 0.50 mg and 0.61 mg, respectively, by analytical g.l.c.

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